NOVEL PROCOAGULANT PROTEINS

This invention relates to a novel series of proteins which exhibit procoagulant properties. These proteins have marked structural differences from human factor VIII:C, but have 5 similar procoagulant activity.

Factor VIII:C is the blood plasma protein that is defective or absent in Hemophilia A disease. This disease is a hereditary bleeding disorder affecting approximately one in 20,000 males. 10 The structure of factor VIII:C is described in U.S. Patent Applications Serial No. 546,650 filed october 28, 1983 and No. 644,036 filed August 24, 1984, which are incorporated herein by reference and in Nature, 312:306, 307, 326 and 342.

15 One of the problems presently encountered with the use of human factor VIII:C for treatment of hemophilia arises from its antigenicity. A significant percentage of hemophiliacs have developed an immune reaction to the factor VIII:C used for their treatment. Non-hemophiliacs can also develop or acquire $_{
m 20}$ hemophilia when their immune systems become sensitized to factor VIII:C and produce circulating antibodies or "inhibitors" to factor VIII:C. In either case, the effect is the neutralization of whatever factor VIII:C is present in the patient, making treatment very difficult. Until now, the method of 25 choice for treating hemophiliacs with this problem has been to administer, in cases of severe bleeding episodes, non-human factor VIII:C, such as treated porcine factor VIII:C. Kernoff et al., <u>Blood</u> 63:31 (1984). However, the antibodies which neutralize the clotting ability of human factor VIII:C 30 will react to a varying extent with factor VIII:C of other species, and the porcine protein is itself antigenic, thus both the short-term and long-term effectiveness of such treatment will vary.

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Additionally, patients frequently display adverse reactions to infusion with the porcine factor VIII:C. The use of porcine factor VIII:C in spite of the risks has been justified because of the lack of reliably effective alternatives. Kernoff, supra at 38. The present invention provides an alternative to the administration of porcine factor VIII:C.

This invention provides for proteins which have procoagulant activity similar to that of factor VIII:C and also have substant10 ially lower molecular weight. These proteins are schematically depicted by formula (1) as follows:

 f_{15} wherein A represents a polypeptide sequence substantially duplicative of the sequence Ala-20 through Arg-759; B represents a polypeptide sequence substantially duplicative of the sequence Ser-1709 through the C-terminal Tyr-2351; and X represents a polypeptide sequence of up to 949 amino acids substantially 20 duplicative of sequences of amino acids within the sequence Ser-760 through Arg-1708. The amino terminus of region X is covalently bonded through a peptide bond (designated "-" in formula 1) to the carboxy terminus of A. The carboxy terminus of region X is likewise bonded to the amino terminus of B. 25 Numbering of amino acids throughout this disclosure is with reference to the numbering of amino acids in Table 1 in which the first amino acid, Met, of the leader sequence is assigned Protein domain X may comprise a continuous but shorter sequence selected from the region Ser-760 through 30 Arg-1708. Alternatively X may comprise two or more amino acid sequences selected from that region which are covalently bonded by a peptide bond (maintaining an ascending numerical order of amino acids).

By way of example, one compound of this invention contains a region X comprising the amino acid sequence of Ser-760 to Pro-

T0040X

TABLE 1

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TTACTTTTTTCCCCTCCTGCGGGCCTAAAGATATTTTAGAGAAGAATTAACCTTTTGCTTCTCCAGTTGAACATTTGTAGCAATAAGTC HET CIN Ile Clu Leu Ser Thr Cys Phe Phe Leu Cys Leu Leu Arg Phe Cys Phe ATG CAA ATA GAG CTC TCC ACC TGC TTC TTT CTG TGC CTT TTG CGA TTC TCC TTT Ala Thr Λrg Cly Clu Arg Leu Val Tyr Tyr Ala Len Ser Trp Asp Tyr RET TAC CTC ACT AGA AGA TAC CGT CCV CTC CAA CTC TCA TCC Clu Cly Ala Cln Ser Pro Val Phe Asp Leu Leu Asp Arg Pro Pro Arg Val Pro CAA AGT GAT CTC GGT CAG CIC CCT CTC CAC CCA ACA TIT CCT CCT AGA Ser Phe Pro Phe Lys Asn Thr Ser Val Val Tyr Lys Lys Thr Leu Phe Val TCA CCA AAC ACC CTC CTC TAC AAA AAG ACT CTG TIT CTA Val His Phe Asn He Ala Lys Pro Arg Pro Pro Trp HET Gly Leu CAC CIT TTC AAC ATC CCT AAC CCA ACC CCA CCC TCC ATC CCT CTC Lys 108 Gin Glu Va1 Tyr Thr .Val Val ASP Ile Thr Læu ACC ATC CAG CCT CGT CCT CAG CTT CTA TAT CAT ACA CTC CTC ATT ACA CTT AAG Trp Lys 126 Ser Pro Val Ser Leu His Alá Val Cly Val Ser Tyr CCT GTC ACT CAT TCC CTI CAT CCT GTT CCT GIA TCC TAC TGG AAA Glu Asp 144 Cly Ala Clu Cln Arg Tyr Asp ASD Gin The Ser Clu Ly s GGA GCT GAA TAT GAT CAT CAG ACC ACT CAA ACC CAC MA GAA CAT Lys Glu 162 Pro Gly Gly Ser His The Va1 lyr Ttp C1n Val Leu CCA AAA TIC CCT CCT ACC CAT ACA TAT CTC TCC CAC CTC CIC AAA GAG Ser His 180 Pro Ser Asp Leu Cys Leu Thr Tyr Ser Tyr Leu AAT CCT CCA ATC CCC TCT CAC CCA CIC TCC CTT ACC TAC TCA TAT CTT TCT CAT Val Lys ASD Leu Asp Ser Cly Leu A3n Leu Ile Gly Ala Val Cya 198 Leu Leu GAC CTG CTA AAA GAC TTG AAT TCA CCC CTC ATT GGA CCC CTA CTA CTA TGT Cly Ser Leu Clu Ala Lys Lys The Gln The Leu H1s Lys Phe Tie Leu 216 AGT CTG CCC AAG CAA AAG ACA CAC TTC CAC ACC ** III ATA CTA Phe Ala Val Cly Ser 234 Asp Glu Lys Ser Clu Thr Trp His Ser LYS Asn CTT TIT CCT CTA TTT CAT CAA CCC ACT TCC CAC TCA GAA ACA AAG AAC TCC Cln ABD Asp Ala Thr 252 Arg Ala Ser Ala Arg Pro MET HIS Ala Trp Lys CAT TIC ATG CAG CAT CCT ACC CCA TCT CCT CCC TCC CCT AAA ATC CAC ACA Tyr Val Cty Lys 270 Asn Ser Pro Clv Arg Leu Leu Lic Cys His CTC CTA CCT TAT AAT AAC ACC ICI CTG CCA CCT CTG ATT CÇA TCC CAC ACC His Val Cly Cly Ilc 288 Scr Tyr Trp Ile MET The Thr Pro Clu Val His Ser CTC TAT TCC CAT CTC ATT GCA ATG CGC ACC ACT CCT CAY GTC CAC TCA Asn His Arg Cln Ala Ser Leu Glu AAC CAT CGC CAC CCG TCC TTG GAA Clv His Phe Val Clu 306 Thr Leu Arg ITC CTC GAA CCT CAC ACA TTT CTT CTG ACC

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Ile ATC	Ser TCG	Pro	Ile ATA	Thr ACT	Phe TTC	l.eu CTT	The	Ala CCT			Leu CTC	Lcu TTC	MET ATC	Asp CAC	Leu CTT	Gly	Gln CAG	324
		Leu	Phe	Сув	llis	Ile	Ser	Ser	His	Gin	His	Λsp	Gly	MET	Clu	۸la	Tyr	342
III	CTA	CTG	III	TCT	CAT	ATC	TCT	TCC	CAC	CVV	CAT	GAT	CCC	ATC	GAA	CCT	TAT	
Val GTC	-,-	Val CTA	Asp GAC	Ser AGC	Cys TCT		Glu GAG	Clu CAA	Pro CCC	G1n	l.eu CTA	Arg CGA		Lys AAA	۸sn ۸۸.۲	Asn AAT	Clu CAA	360
C1u		Clu	Asp	Tyr	Asp	•	Asp			•		Clu		Λsp	Val	Val	Arg	378
GAA	CCC	CAA	GAC	TAT	CAT	CAT		CTT			TCT	CVV	ATG	GAT	GTG	CTC	AGG	396
Phe TTI	Asp GAT	Asp GAT	Asp GAC	Asn AAC	Ser ICI	Pro	Ser TCC	Phe III		Cln CAA	Ile ATT	Arg CCC	Ser TCA	Val CTT	Ala GCC	Lys	Lys	
His CAT	Pro	Lys	Thr	Trp	Val GTA	His CAT	Tyr		Aln GCT		Glu	Glu G1G	Glu GAG	Λsp GAC	Irp	Λsp	Tyr	414
Ala	Pro	Leu	Val	Leu	Ala	Pro	Asp	Asp		Ser	GAA	CAC Lys	Ser		TCC	GAC	TAT	432
CCT	ccc	ITA	CTC	CTC	GCC	ccc	CAT	•	AGA		TAT	AAA	ACT	CAA	Tyr TAT	Leu TTG	AAC	
Asn AAT	Gly GCC	Pro	Gln CAG	Arg CGG	Ile ATT	Gly GGT	Arg AGG	Lyn AAC	Tyr TAC	Lys	Lys	Val UTC	Arg CCA	Phe	HET ATG	Ala CCA	Tyr	450
Thr		Clu	Thr	Phe	Lys	Thr	Arg			lle	Cln	His	Glu	Ser	Gly	Ila	Leu	468
ACA	CAT	GAA	ACC	111	AÁC	ACT	CCT		CCT	ATT	CAG	CAT	CAA	TCA	CCV	ATC	TTG	
Gly GGA	Pro CCT	Ľeu TTA	Leu CTT	Tyr TAT	Gly GGC	Glu GAA	Val GTT		GAC GAC	Thr ACA	Leu CTG	Leu TTC	Ile ATT	Ile ATA	Phe TTT	Lys AAG	Asn AAT	486
	Ala		-	Pro	Tyr		Ile			His	•	He	Thr	Asp	Val	ĀTģ	Pro	504
	CCA	AGC		CCA	TAT	AAC	ATC	TAC	CCT	CAC	CGA	ATC	ACT	CAT	CTC	CCT	CCT	533
Leu TTC	TAT	TCA	Arg AGG	ACA	Leu TTA		Lys	Gly	CTA	Lys	CAT	Leu TTG	Lys	CAT	Phe III	Pro	Ile ATT	522
Leu CTG		Cly GGA			Phe TTC		Tyr TAT	Ly-s AAA	Trp TGG	Thr ACA	Val GTG	Thr	Val GTA	Glu GAA	Asp GAT	Gly GGG	Pro CCA	540
Thr	Lys		Asp		Arg		Leu			Tyr	Tyr	Ser		Phe	Val	Asn	HET	558
ACT				CCT	ccc	TGC	CTC	ACC	•	TAT	TAC	TCT	AGT	TTC	CII	AAT	ATG	•
Glu CAG	Arg AGA		Leu CTA			Gly GGA			Gly GGC	Pro CCT	l.eu CTC	Leu CTC	lle ATC	Cys TCC	Tyr TAC	Lys AAA	Glu GAA	576
	Val				Cly	Asn		He	HLT	Ser	Азр	Lys	·Λrg	Asn	Val	lle	Leu	594
TCT	CTA			ACA		VIC			ATG	TCA	GAC	AAG	VCC	TAA	GTC	ATC	CTC	
Phe	Ser	Vn1 CTA	Phe TTT	Asp CAT		AAC	VCV	VCC Set	Trp	Tyr	Leu CTC	The ACA	Glu GAG	Asn AAT	lle ATA	Cln CAA	Arg CGC	612
Phe TTT	Leu CTC	Pro	Asn AAT			Cly					۸sp	Pro	Clu	Plie	Gln	۸la	Ser	630
Asn	Ile	MET	His	Ser		GGA Asn			CTT		GAT	CCA	GAG	TTC	CAA	CCC	TCC	648
AAC	ATC		CAC	ACC	ATC	AAT	CCC	ZVI	CTT	Phe TTT	CAT	Ser	Leu IIC		Leu TTC	Ser TCA	U _e j GTT	040

Cy* TGT	Leu TTC	His	CAG	Val CTG	Ala CCA	Tyr	Trp	Tyr	Ile ATT	Leu CTA	Ser ACC	Ile ATT	Cly CCA	Ala GCA	GIn CAG	Thr	Λsp C AC	666
	. Leu CTT			Phe TTC	Phe TTC	Ser TCT	Gly CGA	Tyr	Thr	Phe TTC	Lys AAA	His CAC	Lys AAA	MET ATG	Val GTC	Tyr TAT	Glu CAA	684
Asp GAC	Thr ACA	Leu CTC	Thr ACC	Leu CTA		Pro CCA		Ser TCA					Phe TTC	MET ATC	Ser TCG	MET ATC	Clu CAA	702
			Leu CTA		Ile ATT	Leu CTG	G1y GCG	Cy's TGC	His CAC	Asn AAC	Ser TCA	Asp CAC	Phe TTT	Arg CCG	Asn AAC		Gly CCC	720
HET ATC	Thr ACC	Ala CCC	Leu TTA	Leu CTG	Lys AAC	Val GTI	Ser TCT	Ser AGT	Cys IGT	A#P GAC	Lys AAG	Asn AAC	Thr ACT	Cly GGT	Asp GAT	Tyr TAT	Tyr TAC	738
	Asp GAC		Tyr TAT		Asp CAT	Ile AIT	Ser TCA	Ala GCA	Tyr TAC	Leu TTC	l.eu CTG	Ser AGT	Lys AAA		Asn AAT	Ala GCC	Ile ATT	756
Glu GAA	Pro CCA	Arg AGA	Ser ACC	Phe TTC	Ser TCC	Gln CAG	A&II.	Ser TCA	Arg	His CAC	Pro CCT	Ser AGC	Thr	_		-	Gln CAA	774
Phe TIT	Asn		Thr	Thr ACA	lle ATT	Pro CCA	Clu CAA	Asn AAT	Asp GAC	lle ATA	Glu CAG	Lys AAG	Thr ACT	Asp GAC	Pro CCT	•	Phe TTT	792
Ala GCA	His CAC	Arg AGA	Thr	Pre CCT	MET ATG	Pro CCT	Lys AAA	Ile ATA	Gln CAA	nzA TAA	Val CTC	Ser TCC	Ser TCT	Ser ACT	Asp CAT	Leu TTC	Leu TTC	810
MET ATG	Leu CTC	Leu TTC	Arg	Gln CAG	Ser ACT	Pro CCT	Thr ACT	Pro CCA	His CAT	Gly CCG	Leu CTA	Ser TCC			Asp GAT		Cln CAA	828
Glu GAA	Ala GCC	Lys	Tyr TAT	Clu GAG	Thr ACT	Phe	Ser TCT	Asp CAT	Asp GAT	Pro	Ser TCA	Pro CCT	Gly GGA	Ala CCA	Ile ATA	qsA CAC	Ser ACT	846
Asn TAA	Asn AAC	Ser AGC	Leu CTG	Ser TCT	Glu GAA	MET ATC	Thr	CVC	Phe TTC	Arg ACC	Pro CCA	Cln CAC	Leu CTC	His CAT	His CAC		Gly CGG	864
Asp GAC	MET ATC	Val CTA	Phe TTT	Thr ACC	Pro CCT	Clu GAG	Ser TCA	CCC	Leu CTC	Cln CAA	Leu TTA	ATR	Leu TTA	Asn AAT	Glu GAG			882
Cly CCC	Thr ACA	Thr ACT	Ala GCA	Ala GCA	Thr	Glu GAG	Leu TTG	Lys AAG	Lys AAA	Leu	Asp GAT	The TTC		Val CTT	Ser TCT	Ser ACT	Thr	900
Ser TCA	Asn AAT	neA TAA	Leu CTG	Ile ATT	Ser TCA	Thr	Ile ATT	CCA		Asp CAC				Ala GCA		The ACT	Amp GAT	918
Asn AAT	Thr ACA	Ser AGT	Ser TCC	Leu TTA	Gly	Pro	Pro	Ser ACT	MET ATG	Pro	Val CTT	His CAT	Tyr TAT	Asp GAT	Ser ACT	Cln CAA	Leu TTA	936
Asp CAT	Thr		Leu CTA	Phe ITT	Gly	Lys	Lys AAC	Ser TCA	Ser TCT	Pro CCC	t.eu CTT		Clu GAG		CLY			954
Leu CTC		Leu TTC	Ser ACT	Glu CAA	Clu CAA	naA TAA		Asp CAT			Leu TTG		Clu CAA	Ser TCA	G1y GCT			972
neA TAA	Ser ACC	Gln CAA	Glu CAA	Ser ACT	Ser TCA	Trp TGG	Gly CCA	Lye	Asn AAT	Val CTA	Ser TCG	Ser TCA	Thr	Glu GAG	Ser AGT	Cly	Arg	990

Leu TIA	Phe TTT	Lys AAA	Gly CGG	Lys	Arg AGA	Ala GCT	H1s CAT	G1y GCA	Pro CCT	Ala GCT	Len TTG	I.eu TTG	Thr ACT	Lys Aaa	Amp GAT	Λsπ ΛΛΙ	GCC
Leu TTA	Phe TTC	Lys AAA	Val CTI	Ser AGC	Ile ATC	Ser TCT	Leu	Leu TTA	Lys AAG	Thr ACA	Λε·π AΛC	Lys AAA	Thr	Ser TCC	ASN AAT	Asn AAT	Ser 1,026 TCA
Ala CCA	Thr ACT	neA TAA	Arg ACA	Lys AAC	Thr	H1s CAC	Ile ATT	Asp CAT	Cly CCC	Pro CCA	Ser TCA	Leu TTA	Leu TTA	Ile ATT	Glu GAG	Asn AAT	Ser 1,044 AGT
Pro CCA	Ser TCA	Val GTC	Trp	Gln CAA	Asn AAT	ile ATA	Leu TTA	Glu GAA	Ser AGT	Asp GAC	Thr ACT	Glu CAG	Phe TTT	Lys AAA	Lys AAA	Val CTC	Thr 1,062 ACA
Pro CCT	Leu TTG	Ile ATT	His CAT	Asp GAC	Arg AGA	MET	Leu CTT	HET ATG	Asp GAC	Lys	Asn TAA	Ala GCT	Thr	Ala CCT	Leu TTC	ACC	Leu 1,080 CTA
Asn TAA	K1s CAT	HET ATC	Ser TCA	Asn AAT	Lys AAA	Thr ACT	Thr ACT	Ser TCA	Ser TCA	Lys	Λ s n ΛCC	MET ATG	Glu GAA	MET ATC	Val CTC	Cln CAA	Cln 1,098 CAG
Lys AAA	Lys AAA	Glu GAG	Gly GGC	Pro CCC	Ile ATT	Pro CCA	Pro CCA	Asp CAT	Ala CCĄ	Gln CAA	nsh TAA	Pro CCA	Asp GAT	HET ATG	Ser 1CG	Phe TTC	Phe 1,116
Lys AAG	MET ATC	Leu CIA	Plie TTC	Leu TTG	Pro CCA	Glu GAA	Ser TCA	Ala CCA	Arg ACC	Trp	Tle ATA	Gln CAA	Arg ACG	Thr ACT	H1s CAT	Cly	Lys 1,134 AAG
Asn AAC	Ser TCT	Leu CTG	AAC OAA	Ser TCT	Cly CCG	CAA	Cly GGC	Pro CCC	Ser ACT	Pro CCA	Lys AAC	Cln CAA	Lcu TTA	Val GTA	Ser TCC	Leu TTA	Cly 1,152 CCA
Pro	Clu GAA	Lys AAA	Ser TCT	Val GTG	Clu GAA	Gly	Gln CAG	nsA TAA	Phe TTC	Leu TTC	Ser TCT	G) u GAG	Lys AAA	Asn A.C	Lys AAA	Val CTC	Val 1,170 CTA
Val GTA	G17 GCA	Lys AAC	Gly	G1u GAA	Flie TTT	Thr ACA	Lys AAG	Asp CAC	Val GTA	CCA	Len	Lys AAA	Glu CAG	HET ATG	Val CTT	Phe ITT	Pro 1,188 CCA
Ser AGC	Ser AGC	Arg AGA	Asn	Leu CTA	Phe TTT	Leu CTT	Thr	Asn AAC	Leu TTC	Asp GAT	ncA TAA	Leu TTA	His CAT	Glu GAA	Asn AAT	nzA TAA	Thr 1,206
H1s CAC	Asn TAA	C1n CAA	Clu CAA	Lys AAA	Lys	Ile ATT	C1n CAC	Clu CAA	G1u CAA	lle ATA	Glu GAA	Lys AAC	Lys AAG	Glu GAA	Thr	Leu TTA	Ile 1,224 ATC
Gln CAA	Glu GAG	Asn AAT	Val.	Val CTT	Leu TTC	Pro CCT	C1n CAG	Ile	H1s CAT	Thr	Val CTC	Thr	CCC	Thr ACT	Lys AAG	Asn ANT	Phe 1,242
MET ATC	Lys	AAC	Leu CIT	Phe TTC	Leu TAA	Leu CTG	Ser	Thr	VCC VCC	CIA CAA	Asn AAT	Val GTA	Glu GAA	Gly GGT	Ser TCA	Tyr TAT	Glu 1,260 GAC
GIy GGG	Ala GCA	Tyr	Ala CCT	Pro	Val CTA	Leu CTT	Gln CAA	CAT	Phe TTT	Arg ACC	Ser TGA	Leu	neA TAA	Asp CAT	Ser TCA	Thr	Asn 1,278 AAT
ACA	Thr	Lys	Lys	CVC	Thr	Ala CCT	His CAT	Phe	Ser TCA	Lys	Lys	Gly	G1u CAG	CAA	CVV CVV	AAC	Leu 1,296
Glu GAA	CCC	Leu TTC	CCA	Asn TAA	Cln CAA	Thr	Lys AAC	CAA	lle ATT	Va I CTA	Glu GAG	Lys	Tyr TAT	VI#	Cys	Thr	Thr 1,314
ACC	Ile ATA	Ser TCT	Pro CCT	Asn AAT	Thr	Ser AGC	Gln CAG	C1n CAG	ASN AAT	Phe TIT	Val CTC	Thr	CAA	Arg CCT	Ser ACT	Lys AAC	Arg 1,332 AGA



Ala CCT	Leu TTC	Lys AAA	CAA	Phe TTC	Arr Aga	Leu CTC	Pro CCA	Leu CTA	Glu GAA	C.L.\	Thr	Clu CAA	Leu CTT	Glu GAA	Lys AAA	Arg AGC		1,350
Ile'	Val GTG	Λsp GAT	Asp GAC	Thr ACC		Thr	Cln CAC	Trp	Ser TCC	Lys AU	Asn AAC	MET ATC	Lys MA	His CAT	l.cu TTC	Thr	Pro-	1,368
Ser AGC	Thr	Leu CTC	Thr ACA	G]n CAG	Ilc ATA	Asp GAC	Tyr TAC		Glu CAC			Lys AAA			Ile ATT	Thr ACT	Cln CAC	1,386
Ser TCT	Pro CCC	Leu TTA			Cys TGC	Leu CTT			Ser ACT				Pro CCT		Ala GCA	Asn AAT		1,404
Ser TCT	Pro	Leu TTA	Pro	Ile ATT	Ala GCA	Lys AAG	Val GTA	Ser TCA	Ser TCA	Pha TTT	Pro CCA	Ser TCT	Ilo ATT		Pro CCT		Tyr TAT	1,422
Leu CTG	Thr	AFG	Val GTC		Phe TTC		Asp GAC		Ser TCT				Pro CCA			Ser TCT		1,440
ACA		Lys	Asp GAT	Ser TCT	Cly CGG		Cln CAA		Ser ACC		His					Ala GCC		1,458
Eys AAA	ns/. TW.	Asn	Leu CTT	Ser TCT	Leu TTA	Ala GCC	Ile ATT		Thr			MET ATC	Thr			CIn CAA		1,476
Glu GAG	Val CTT	Gly	Ser TCC	Leu CTG	Gly GCG	Thr ACA	Ser AGT	Ala GCC	Thr				Thr ACA	Tyr TAC		Lys	Val GTT	1,494
CAC	Asn AAC	Thr	Val GTT	Leu CTC	Pro CCG	Lys	Pro CCA	Asp GAC		Pro CCC			Ser TCT	Gly CGC	Lye		Glu GAA	1,512
Leu TTC	Leu CTT	Pro	Lys	Val CTT	His CAC	Ile ATT	Tyr TAT	Gln CAG	Lys AAC	CVC YRb	Leu CTA	Phe TTC	Pro CCT	Thr	Glu GAA		Ser AGC	1,530
AAT	CCC	TCT		CCC	CAT	CIG	CAT	CTC	CTC	CAA	GCC	ACC	CTT	CII	Gln CAG	Gly GGA	Thr	1,548
Glu GAG	Cly CCA	Ala GCG	Ile ATT	Lys AAC	Trp TGG	Asn AAT	Glu GAA	Ala CCA	Asn AAC	Arg AGA	Pro CCT	Cly	Lys AAA	V#1 CTT	Pro CCC	Phe TTT	Leu CTC	1,566
ACA	Val CTA	Ala CCA	Thr	Glu GAA	Ser	Ser TCT	Ala GCA	Lys AAC	Thr	Pro CCC	Ser TCC		Leu CTA	Leu TTC	Asp GAT	Pro CCT	Leu CTT	1,584
Ala CCT	Trp	Asp Cat	Asn	His CAC	Tyr TAT	Gly	Thr ACT	Gln CAG	Ilc ATA	Pro CCA	Lys AAA	Glu GAA	C1a	Trp TCG	Lys	Ser TCC	Gln CAA	1,602
Glu GAG	Lys AAG	Ser TCA	Pro CCA	Glu GAA	Lys AAA	Thr ACA	Ala GCT	Phe TTT	VVC	Lys	Lys AAC	Asp GAT	The	Ile ATT	Leu TTC	Ser TCC	Leu	1,520
Asn AAC	Ala GCT	Cys TGT	Glu GAA	Ser AGC	Λsn TAA	His CAT	Ala CCA	lle ATA	Ala CCA	Ala CCA	I le ATA	Λ×n TAA	Glu GAG	Gly CCA	Cln CAA	Asn AAT	Lys	1,638
Pro CCC	Clu GAA	Tle ATA	C1u CAA	Val CTC	Thr	Trp	Ala GCA	YVC	Gin CAA	Cly CCT	Arg ACC	Thr	Glu CAA	Arg AGG	Leu CTC	Cys TCC	Ser TCT	1,656
C1n CAA	A&n AAC	Pro CCA	Pro	Vn1 CTC	I.eu TTC	Lys AAA	Arg CCC	H1s CAT	Cin CAA	Arg CGC	Glu CAA	Jle ATA	Thr ACT	Arg CCT	The	The ACT	Len CTT	1,674



Gln CAG	Ser TCA	Asp GAT	Gln CAA	Glu GAG	Glu GAA	lle ATT	Asp UAC	Tyr TAT	Asp CAT	Asp CAT	Thr ACC	ATA	Ser TCA	Val GTT	Glu GAA	MET ATG	Lys AAC	1,692
Lys AAC	Ç1u GAA	Asp GAT	The	Asp CAC	Ile ATT	Tyr TAT	Asp CAT	Glu GAG	Asp CAT	CVV	Asn AAT	G1n CAG	Ser	Pro CCC	Arg CGC	Ser AGC	Phe TTT	1,710
G1n CAA	Lys AAC	Lys	Thr ACA	Arg CCA	His	Tyr TAT	Phe TTT	Ile ATT	Ala GCT	GCA	Val GTC	Clu GAG	ATg AGC	Leu CTC	Trp TCC	Asp Cat	Tyr TAT	1,728
Gly	HET ATC	Ser AGT	Ser AGC	Ser TCC	P T O	His CAT	Val CTT	Leu CTA	Arg	Asn AAC	Arg AGC	Ala CCT	Gln CAG	Ser AGT	Cly GCC	Ser ACT	Val GTC	1,746
Pro CCT	Cln CAG	Phe TIC	Lys AAG	Lys AAA	Val GTI	Val CTT	Phe TTC	Gln CAG	Glu GAA	Phe TTT	Thr	Asp GAT	Gly GGC	Ser TCC	Phe TIT	Thr ACT	G1n CAG	1,764
Pro CCC	Leu TTA	Tyr TAC	Arg CCT	Gly GGA	Glu GAA	Leu CTA	AAT	Glu GAA	His CAT	Leu TTG	Gly GGA	Leu CTC	Leu CTC	Gly GGG	Pro CCA	Tyr	Ile ATA	1,782
Arg AGA	Ala GCA	Glu GAA	Val GII	Clu CAA	Asp CAT	Asn AAT	Ile ATC	MET ATG	Val GTA	Thr ACT	Phe TTC	Arg	Asn AAT	Gln CAG	Ala GCC	Ser TCT	Arg CCT	1,800
Pro CCC	Tyr TAT	Ser TCC	Phe TTC	Tyr TAT	Ser TCT	Ser AGC	Leu CTT	Ile ATT	Ser TCT	Tyr TAT	Glu GAG	Clu CAA	Asp GAT	Gln CAG	Arg AGG	Cln CAA	Gly GGA	1,818
Ala GCA	Glu GAA	Pro CCT	AFB AGA	•	Asn AAC	Phe TTT	Val GTC	Lys AAC	Pro CCT	Asn AAT	Glu GAA	Thr	Lys AAA	The	Tyr TAC	Phe TTT	Trp TGG	1,836
Lys AAA	Val CTC	Gln CAA	His CAT	H1s CAT	MET ATC	Ala GCA	Pro CCC	The	Lys AAA	CVI.	Glu GAG	Phe TTT	Asp GAC	cys TGC	Lys AAA	Ala GCC	Trp	1,854
Ala GCT	Tyr TAT	Phe TTC	Ser TCT	Asp GAT	Val CTT	Asp GAC	Leu CTG	Glu GAA	Lys	Asp CAT	Val GTC	His CAC	Ser TCA	Gly GGC	Leu CTG	Ile ATT	Gly GGA	1,872
Pro CCC	Leu CTT	Leu CTG	Val CTC	Cys TGC	His CAC	Thr	Aen AAC	Thr ACA	Leu CTG	Asn AAC	Pro CCT	Ala CCT	His CAT	Gly CCC	Arg AGA	Cln CAA	Val CTC	1,890
Thr ACA	Val CTA	Glii CAG	Glu GAA	Phe TII	Ala GCT	Leu CTG	Phe TII	Phe TTC	Thr	Ile ATC	Phe TTT	Λsp CAT	Glu GAG	Thr	Lys AAA	Ser AGC	Trp	1,908
Thy TAC	Phe TTC	Thr ACT	Clu CAA	Asn AAT	MET ATC	Clu CAA	Arg AGA	Asn AAC	Cys TGC	Arg AGC	Ala	Pro CCC	Cys TGC	Asu	Ile ATC	Gln CAG	MET ATC	1,926
Clu Caa	Asp CAT	Pro CCC	Thr ACT	Phe TTT	Lys AAA	Glu CAG	Asn AAT	Thr TAT	Arg CGC	Plie TTC	His CAT	Ala GCA	Ile ATC	ns\ TAA	Gly GGC	Tyr TAC	ile ATA	1,944
HET ATG	Asp CAT	Thr	Leu CTA	Pro CCT	Cly CCC	Leu TTA	Val CTA	MET AIG	Ala GCT	Gln CAG	Asp CAT	Gln CAA	Arg AGG	Ile ATT	Arg CGA	Trp	Tyr TAT	1,962
Leu CTG	Leu CTC	Ser AGC	HET ATC	Cly CCC	Ser AGC	AAT	Glu CAA	Asn AAC	11e ATC	IIIs CAT	Ser TCT	Ile ATT	H1s CAT	Phe TTC	Sec AGT	Cly GCA	His CAT	1,980
Val CTG	Phe TTC	Thr ACT	Val CTA	Arg CCA	Lys AAA	Lys AAA	Glu CAG	Glu GAG	Tyr 1AT	Lys A/A	MET ATC	Ala GCA	1.eu CTG	Tyr TAC	Asn AAT	1.eu CTC	Tyr	1,998
Pro CCA	Cly	Val CTT	Phe TTT	Clu CAC	Thr	Val GIG	Glu GAA	MET ATG	Leu TTA	OT9 AUQ	Ser ICC	Lys AAA	Ala CCT	Cly CEA	Ile ATT	Trp TCC	Arg CGG	2,016

	TAI	3LE	그, (cont	inu	ed												
			Leu CTT														Leu CTG	2,034
Val GTG		Ser AGC	Asn AAT	Lys AAG				Pro CCC					Ser TCT				Arg AGA	2,052
-	Phe TTT							Gln CAA									Ala GCC	2,070
Arg ACA	Leu CTI		Tyr TAT												P TO CCC	Phe	Ser TCT	2,088
			Val CTG					Pro CCA									Gln CAG	2,106
Cly CCT	Ala GCC	Arg CGT	Gln CAG	Lys AAG	Phe TTC	Ser TCC	Ser AGC	Leu CTC	Tyr TAC	Ile ATC	Ser TCT	Gln CAG	Phe TTT		Ile ATC	MET ATG	Tyr TAT	2,124
Ser AGT	i.eu CTT	Asp GAT	Gly GGG	Lys AAG	Lys AAG	Trp TGG	Gln CAG	Thr ACT	Tyr TAT	Arg CGA	Gly GCA	Aen AAT	Ser TCC		Gly CCA		Leu TTA	2,142
MET ATC	Val CTC	Phe TTC	Phe TIT	Gly GGC	Asn TAA	Val CTC	Asp CAT	Ser TCA	Ser TCT	G1y CCC	Ile ATA	Lys AAA	His CAC	Asn AAT	Ile ATT	Phe TTT	Asn AAC	2,160
Pro CCI	Pro ADD	Ile ATT	Ile ATT	Ala	Arg CGA	Tyr	Ile ATC	Arg CCT	Leu TTC	H1s CAC	Pro CCA	Thr	H1s CAT	Tyr TAT		lle ATT	Arg CCC	2,178
Ser AGC	Thr ACT	Leu CTT	Arg CGC	MET ATG	Glu GAG	Leu TTG	MET ATG	Cly CCC	Cys TCT	Asp CAT	Lru TTA	neΛ	Ser AGT	Cys TCC	Ser ACC	HET	Pro CCA	2,196
Leu TTG	Gly CCA	MET ATC	Glu GAG	Ser ACT	Lys AAA	Ala GCA	Ile ATA	Ser TCA	Asp GAT	Ala GCA	C1n CAG	lle	Tlir ACT		Ser TCA		Tyr TAC	2,214
Phe TTT	Thr	Asn	HET ATG	Phe	Ala GCC	Thr	Trp TGG	Ser TCT	Pro CCT	Ser TCA	Lys AAA	Ala GCT	Arg CGA			Leu CTC	Gln CAA	2,232
Cly	Arg AGG	Ser ACT	Asn AAT	Ala	Trp TGG	Arg AGA	Pro CCT	Gln CAG	Val GTC	ne/. TA/.	Asn AAT	Pro CCA	Lys AAA	Glu CAG	Trp TCG	Leu CTC	Gln CAA	2,250
Val CTC	Asp GAC	Phe TTC	Gln CAG	Lys AAC	Thr	HET ATG	Lys AAA	Val CTC	Thr ACA	Gly GGA	Val CTA	Thr ACT	The ACT	Gln CAG	Gly GCA	Val CTA	Lys AAA	2,268
	Leu CTG			Ser	MET ATC	Tyr TAT	Val CTC	Lys AAC	Glu GAG	Phe TTC	Leu CTC	Ile ATC	Ser TCC	Ser	Ser ACT	CAA	Asp CAT	2,286
	HIS CAT			Thr ACT	Leu CTC	Phe TTT	Phe TIT	C1n CAG	Asn AAT	Cly CCC	Lys	Val CTA	Lys AAC	VAL	Plie TTT	Cln CAG		2,304
AAT	Cln CAA	Asp GAC	Ser TCC	Phe TTC	Thr	Pro CCT	Val CTC	Val CTC	Asn AAC	Ser TCT	Leu CTA	(:VC	Pro CCA	Pro	Leu TTA	Leu CTG	Thr	2,322
CCC		CTT	CCY	ATT	CAC	CCC	CAG		TCC	CIÇ	CVC	C.\G	Ile ATT	Ala	Leu CTG	Arg ACC	MET ATG	2,340
CAG	CTT	CTG	GCC	TGC	CAG	GCA	CAG	GAC	CIC	TAC	TGA							2,352
			TCCTC/ AACTAI															
															. vvc 1 (LIACO	LIAII	

1000 followed by the amino acid sequence of Asp-1582 to Arg1708. That compound thus comprises the polypeptide sequence of Ala-20 to Pro-1000 covalently linked by a peptide bond to amino acids Asp-1582 to Tyr-2351. Another exemplary compound 5 contains a region X comprising the amino acid sequence Ser-760 to Thr-778 followed by the sequence Pro-1659 to Arg-1708. That compound thus comprises the polypeptide sequence Ala-20 to Thr-778 covalently linked by a peptide bond to the sequence Pro-1659 through Tyr-2351. Still another exemplary compound 10 contains a region X comprising the amino acid sequence Ser-760 to Thr-778 followed by the sequence Glu-1694 to Arg-1708. That compound thus comprises the polypeptide sequence Ala-20 to Thr-778 covalently linked by a peptide bond to amino acids Glu-1694 through Tyr-2351.

15 These exemplary compounds are depicted schematically in Table 2.

The amino acid sequence represented by X should be selected so that it does not substantially reduce the procoagulant 20 activity of the molecule, which activity can be conveniently assayed by conventional methods. Compound (2) of Table 2 is a presently preferred embodiment.

The procoagulant protein may be produced by appropriate host cells transformed by factor VIII:C DNA which has been specifically altered by use of any of a variety of site-specific mutagenesis techniques which will be familiar to those of ordinary skill in the art of recombinant DNA.

30 The starting materials may be a DNA sequence which codes for the complete factor VIII:C molecule, e.g., the complete human factor VIII:C as shown in Table 1, a truncated version of that sequence, or it may comprise segments of that DNA sequence, so long as the starting materials contain at least sufficient DNA to code for the amino acid sequences of the desired polypeptide.



acids deleted relative to human factor VIII:c.

numbering of the sequence depicted in Table 1; and "deletion" indicates the number of amino

0120X

	Compound	Amino Acid Sequence	×	Deletion
	(human factor VIII:c)	(Ala ₂₀ >Tyr ₂₃₅₁₎	(Ser ₇₆₀ ->Arg ₁₇₀₈)	0
	H	$(Ala_{20} \rightarrow Pro_{1000}) - (Asp_{1582} \rightarrow Tyr_{2351})$	$(Ser_{760} \rightarrow Pro_{1000}) - (Asp_{1582} \rightarrow Arg_{1708})$	581
	2	(Ala ₂₀ ->Thr ₇₇₈) -(Pro ₁₆₅₉ ->Tyr ₂₃₅₁₎	$(Ser_{760} \rightarrow Thr_{778}) - (Pro_{1659} \rightarrow Arg_{1708})$	880
-11-	ω	(Ala ₂₀ ->Thr ₇₇₈) -(Glu ₁₆₉₄ ->Tyr ₂₃₅₁)	(Ser ₇₆₀ ->Thr ₇₇₈) -(Glu ₁₆₉₄ ->Arg ₁₇₀₈)	915
	A and B sequence	A and B are as defined, supra; "-" represents a peptide bond; "->" indicates a possequence inclusive of the specified amino acids; amino acid numbering corresponds are as fall the security of the security o	eptide bond; "->" indicates a polypeptide amino acid numbering corresponds to the	g e

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The proceagulent proteins of the present invention, in addition to lacking a substantial amino acid segment of human factor VIII:C, also have fewer potential N-glycosylation sites than human factor VIII. Preferably, at least one N-glycosylation site 5 has been deleted. More preferably, 18 of the 25 potential N-glycosylation sites are not in the molecule. In still more preferred embodiments, up to 19 of the 25 potential N-glycosylation sites are removed. While not wishing to be bound by theory, it is presently believed that the antibodies to factor 10 VIII:C which are directed to antigenic determinants contained in the protein segment deleted in accordance with this invention, i.e., in the amino acid segement itself or in the carbohydrate portion of the glycosylated protein, will not neutralize the procoagulant proteins of the present invention. 15 the fact that the procoagulants of the present invention lack many of the sites for non-human glycosylation by the non-human mammalian or other cells used to produce the proteins is also belived to reduce the antigenicity of that protein, and lessen the likelihood of developing antibodies to the procoagulants. 20 This may enable facilitating the treatment of patients in need of procoagulant therapy.

I contemplate that my compounds can be produced by recombinant DNA techniques at a much lower cost than is possible for pro25 duction of human factor VIII. The host organisms should more efficiently process and express the substantially simpler molecules of this invention.

The compounds of this invention can be formulated into pharmaceu-30 tically acceptable preparations with parenterally acceptable vehicles and excipients in accordance with procedures known in the art.

The pharmaceutical preparations of this invention, suitable for 35 parenteral administration, may conveniently comprise a sterile lyophilized preparation of the protein which may be reconsti-

tuted by addition of sterile solution to produce solutions preferably isotonic with the blood of the recipient. The preparation may be presented in unit or multi-dose containers, e.g. in sealed ampoules or vials. Their use would be analogous to that of human factor VIII, appropriately adjusted for potency.

One method by which these proteins can be expressed is by use of DNA which is prepared by cutting a full-length factor VIII:C DNA with the appropriate restriction enzymes to remove a portion of the DNA sequence that codes for amino acids 760 to 1708 of human factor VIII:C. The cut DNA is then ligated with an oligonucleotide that resects the cut DNA and maintains the correct translational reading frame.

Preparation of the cDNA has been set forth in detail in U.S. Patent Applications Serial Nos. 546,650 and 644,086, supra. A pSP64 recombinant clone containing the nucleotide sequence depicted in Table 1, designated as pSP64-VIII, is on deposit at the American Type Culture Collection under Accession Number ATCC 39812.

Restriction endonucleases are used to obtain cleavage of the human factor VIII: C cDNA, hereinafter the DNA source sequence, at appropriate sites in the nucleotide sequence. 25 otherwise noted, restriction endonucleases are utilized under the conditions and in the manner recommended by their commercial The restriction endonucleases selected herein are those which will enable one to excise with substantial specificity sequences that code for the portion of the factor 30 VIII:C molecule desired to be excised. BamHI and SacI are particularly useful endonucleases. However, the skilled artisan will be able to utilize other restriction endonucleases chosen by conventional selection methods. The number of nucleotides deleted may vary but care should be taken to 35 insure that the reading frame of the ultimate cDNA sequence will not be affected.

The resulting DNA fragments are then purified using conventional techniques such as those set forth in Maniatis et al., Molecular Cloning, A Laboratory Manual (Cold Spring Harbor Laboratory 1982) the disclosure of which is incorporated herein by reference, and 5 Proc. Natl. Acad. Sci. 76:615-619 (1979). The purified DNA is then ligated to form the sequence encoding the polypeptide of the preferred invention. When necessary or desirable, the ligation may be within an oligonucleotide that resects the cut DNA and maintains the correct translational reading frame 10 using standard ligation conditions. Ligation reactions are carried on as described by Maniatis et al., supra at 2453-6 using the buffer described at page 246 thereof and using a DNA concentration of l_{ii} 100 ug/ml, at a temperature of 23°C for blunt ended DNA and 16°C for "sticky ended" DNA. The following 15 double-stranded oligonucleotide is useful when there is BamHI/-SacI deletion such as described infra,

TI 5' P-CATGGACCG-3' PS
TI 3-TCGAGTACCTGGCCTAG 5'; PS

 \sum_{0} but other oligonucleotides can be selected by the skilled artisan depending upon the deletions made and reaction conditions.

The DNA sequences encoding the novel procoagulant polypeptides can, in addition to other methods, be derived from the sequence of human factor VIII:C DNA by application of oligonucleotide-mediated deletion mutagenesis, often referred to as "loopout" mutagenesis, as described for example in Morinaga, Y. et al. Biotechnology, 2: 636-639 (1984).

30 The new DNA sequences containing the various deletions can then be introduced into appropriate vectors for expression in mammalian cells. The procoagulant activity produced by the transiently transfected or stably transformed host cells may 35 be measured by using standard assays for blood plasma samples.



The eukaryotic cell expression vectors described herein may be synthesized by techniques well known to those skilled in this art. The components of the vectors such as the bacterial replicons, selection genes, enhancers, promoters, and the like may be obtained from natural sources or synthesized by known procedures. See Kaufman et al., <u>J. Mol. Biol.</u>, <u>159</u>: 51-521 (1982); Kaufman, <u>Proc. Natl. Acad. Sci. 82</u>: 689-693 (1985).

Established cell lines, including transformed cell lines, are suitable as hosts. Normal diploid cells, cell strains derived from in vitro culture of primary tissue, as well as primary explants (including relatively undifferentiated cells such as haematopoeitic stem cells) are also suitable. Candidate cells need not be genotypically deficient in the selection gene so long as the selection gene is dominantly acting.

The host cells preferably will be established mammalian cell lines. For stable integration of the vector DNA into chromosomal DNA, and for subsequent amplification of the integrated vector DNA, CHO (Chinese hamster ovary) cells are presently preferred. See U.S. Patent 4,399,216. Alternatively, the vector DNA could include all or parts of the bovine papilloma virus genome (Lusky et al., Cell, 36: 391-401 (9184) and be carried in cell lines such as Cl27 mouse cells as a stable episomal element. Other usable mammalian cell lines include HeLa, COS-1 monkey cells, melanoma cell lines such as Bowes cells, mouse L-929 cells, 3T3 lines derived from Swiss, Balb-c or NIH mice, BHK or HaK hamster cells lines and the like.

30 Stable transformants then are screened for expression of the procoagulant product by standard immunological or enzymatic assays. The presence of the DNA encoding the procoagulant proteins may be detected by standard procedures such as Southern blotting. Transient expression of the procoagulant genes during the several days after introduction of the expression





vector DNA into suitable host cells such as COS-1 monkey cells is measured without selection by enzymatic or immunologic assay of the proteins in the culture medium.

The invention will be further understood with reference to the following illustrative embodiments, which are purely exemplary, and should not be taken as limiting the true scope of the present invention, as described in the claims.

DE Cl

EXAMPLE 1

ho 10 ug. of the plasmid pACE, a pSP64 (Promega Biotec, Madison, Wis.) derivative, containing nucleotides 562-7269 of human factor VIII: C cDNA (nucleotide 1 is the A of the ATG initiator meth-5 ionine codon) was subjected to partial BamHI digestion in 100ul containing 50mM Tris. HCl ph 8.0, 50mM MgCl2, and 2.4 units BamHI (New England Biolabs) for 30 minutes at 37°C. reaction was terminated by the addition of EDTA to 20mM and then extracted once with phenol, once with chloroform , ethanol 10 precipitated and pelleted by centrifugation. DNA was redissolved, cleaved to completion in 50ul using 40 units SacI for 1.5 hours at 37°C. DNA was then electrophoresed through a buffered 0.6% agarose gel. An 8.1 kb fragment corresponding to the partial BamHI-SacI fragment of pACE lacking only the 15 sequence corresponding to nucleotides 2992-4774 of the factor VIII:C sequence was purified from the gel using the glass powder technique described in Proc. Nat. Acad. Sci. 76; 615,7619 (1979). Purified DNA was ligated with 100 pmoles of the following double-stranded oligonucleotide

TI 5'P-CATGGACCG-3' PS

TI 3'-TCGAGTACCTGGCCTAG 5' PS

using standard ligation conditions. The DNA sequence removed represents the deletion of 584 amino acid sequence beginning with amino acid 998 and continuing through 1581. The oligonucleotide inserted, however, encodes amino acids corresponding to 998-1000. Therefore, the polypeptide encoded contains deletion of 581 amino acids.

DNA was then used to transform competent <u>E. coli</u> bacteria, and DNA from several ampicillin resistant transformants was analyzed by restriction mapping to identify a plasmid harboring the desired SacI-BamHI deletion mutant. DNA from this plasmid was digested to completion with KpnI, which cleaves the plasmid uniquely at nucleotide 1816 of the factor VIII:C coding se-



quence. This DNA was ligated with a KpnI DNA fragment containing nucleotides 1-1815 of factor VIII:C DNA and a synthetic SalI site at nucleotides 7-11 to -5 and then used to transform competent E. COli bacteria.

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Plasmid DNA was isolated and oriented by restriction mapping to identify a plasmid, pBSdK, containing the correct 5' to 3' orientation of the KpnI insert. SalI digestion, which excises the entire polypeptide coding region from the plasmid, was performed 10 and the DNA electrophoresed through a buffered 0.6% agarose gel. The 5.3Kb SalI fragment was purified from the gel as described above. This DNA fragment was ligated with XhoI cut pXMT2 DNA to give rise to plasmid pDGR-2. pXMT2 is a plasmid capable of expressing heterologous genes when introduced into mammalian 15 cells such as the COS-1 African Green Monkey kidney cell line, and is a derivative of the expression vectors described in Kaufman, supra at 689,793. The expression elements are the same as described for plasmid pQ2 except that it contains a deletion of the adenovirus major late promoter extending from 20 - 45 to +156 with respect to the transcription start site of the adenovirus major late promoter. mRNA expression in pXMT is driven by the SV40 late promoter. The bacterial replicon, however, has been substituted to render bacteria containing the vector resistant to ampicillin rather than tetracycline. 25 pXMT2 contains a unique Xho I site at a position which allows for expression of inserted cDNA from the SV40 late promoter. This Xho I site is convenient for inserting factor VIII:C cDNA constructs since these are flanked by SalI sites.

30 Restriction mapping of transformants identified a plasmid, pDGR-2, containing the correct 5' to 3' orientation of the polypeptide coding sequence relative to the direction of transcription from the SV40 late promoter. pDGR-2 is on deposit at the American Type Culture Collection under Accession number 35 53100.

EXAMPLE 2

CLO

Other novel procoagulant proteins may be obtained from constructs produced by oligonucleotide mediated deletion mutagenesis, using 5 for example the "loopout" mutagenesis techniques as described in Morinaga et al., supra. The deletion mutagenesis is performed using expression plasmid pDGR-2 or any other appropriate plasmid or bacteriophage vector. Other methods for oligonucleotide mediated mutagenesis employing single stranded DNA produced with 10 Ml3 vectors and the like are also suitable. See Zoller et al., Nucl. Acids Res. 10: 6487,7,6500 (1982). For example, these deletions can be produced using the oligonucleotides

AAAAGCAATTTAATGCCACCCACCAGTCTTGAAACGCCA

AAAAGCAATTTAATGCCACCGAAGATTTTGACATTTATGA

f to cause deletions in factor VIII:C cDNA from nucleotides (A) 2334 to 4974 or (B) 2334 to 5079. The proteins encoded by these constructs contain deletions of (A) 880 and (B) 915 amino acids 20 relative to Factor VIII:C.

The deleted constructs are tested directly, or after subcloning into appropriate expression vectors, in order to determine if the novel proteins possess procoagulant activity. Procoagulant 25 activity was assayed as described in Examples 3 and 4.

Expression of Procoagulant Molecules in COS Monkey Cells

30 The expression plasmids containing the modified cDNA's prepared as in Examples 1 or 2 and the full-length cDNA, pXMT-VIII, were introduced into COS-1 cells via the DEAE-dextran transfection protocol. Sompayrac and Dana 1981, Proc. Natl. Acad. <u>Sci</u>. <u>78</u>: 7575,77578. Conditioned media was harvested 48 hours 35 post-transfection and assayed for factor VIII-type activity as described in Toole et. al., 1984, Nature 312:342-347.



results of the experiment are summarized in Table 3. Both plasmids containing the modified cDNAs yielded procoagulant activity and, moreover, the activity was greater than that obtained using wild type cDNA. From these data it was concluded that removal of up to 880 amino acids (95,000 daltons) in a defined domain of human factor VIII does not destroy cofactor activity. Furthermore, these abridged procoagulant proteins retain their ability to be activated by thrombin.

TABLE 3: EXPRESSION OF ABRIDGED FACTOR VIII MOLECULES

5					
		<pre># amino</pre>	chromogenic	Cl	otek
		acids	activity	ac	tivity
	plasmid	deleted	(mUm1 ⁻¹)	-IIa	+IIa (fold)
	No DNA	-	0		•
10					
	IIIV-TMXq	-	15:1	-	450
	pDGR-2	581	114	250	5750 (23X)
15	pLA-2	880	162	. 330	9240 (28X)
		·			

The plasmids indicated were transfected into COS cells and 48 hr. post-transfection the conditioned media taken for assay by the Kabi Coatest factor VIII:C method (chromogenic activity) and by the one-stage activated partial thromboplastin time (APTT) coagulation assay (Clotek activity) using factor VIII:C deficient plasma as described (Toole, Nature 1984). For thrombin (IIa) activation, samples were pretreated 1710 min, with 0.2 units/ml thrombin (IIa) at room temperature. Activation coefficients are provided in parentheses. Activity from media from the wild-type (pXMT-VIII) transfection was too low to directly measure Clotek activity before thrombin activation. From other experiments where the wild type factor VIII activity was concentrated, it was demonstrated to be approximately 30-fold activatable.

EXAMPLE 4

(| Expression of Procoagulant Molecules in CHO Cells

P (A) Expression of pDGR-2

The procoagulant expression vector containing a deletion (relative to the Factor VIII:C cDNA) of 581 amino acids (pDGR-2) was transfected with plasmid pAdD26SV(A) #3 (10 ug pDGR-2:1 ug pAdD26SV(A) #3) by CaPO4 coprecipitation into CHO DHFR deficient cells (DUKX-Bll) and transformants isolated and grown in increasing concentrations of MTX as described by Kaufman et. al., (1985). One transformant designated J1 exhibited the following activities as a function of resistance to increasing concentrations of MTX.

TOZJOX WM MTX	<pre>mUnits/ml/day/10⁶ cells*</pre>
102701 0	1.46
0.02	322
0.1	499
_	

(B) Expression of pLA-2

The procoagulant expression vector containing a deletion of 880 amino acids (pLA-2) was introduced into CHO DHFR deficient cells (DUKX-Bll, Chasin and Urlaub, PNAS 77: 4216-4220, 1980 by protoplast fusion as described (Sandri-Goldin et. al., Mol. Cell. Biol. 1: 743-752). After fusion, fresh medium containing 100 ug/ml of kanamycin, and 10 ug/ml of each of thymidine, adenosine, deoxyadenosine, penicillin, and streptomycin and 10% dialyzed fetal calf serum was added to each plate. The kanamycin was included to prevent the growth of any bacteria which had escaped conversion to protoplasts. Four days later the cells were subcultured 1:15 into alpha-media with 10% dialyzed fetal calf serum, penicillin, and streptomycin, but lacking the nucleosides. Colonies appeared after 10-12 days after subculturing cells into selective media. A group of 8

transformants were pooled and grown in sequentially increasing concentrations of MTX starting at 0.02 uM with steps to 0.1, 0.2, and 1.0 uM MTX. Results of factor VIII-type activity in cells resistant to increasing concentrations of MTX is shown below.

	um mtx	<pre>mUnits/ml/day/10⁶cells*</pre>
T0240X	0	16
	0.02	530
10	0.2	1170
	1.0	1890

* Factor VIII activity was determined by the Kabi Coatest factor VIII: C method (chromogenic activity).

15

B

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